

TUMORS AND THE BRAIN BARRIERS

Co-chairs: Quentin Smith¹, Lauren Abrey²

Contributors: Tracy Batchelor³, Keith Black⁴, Jaishri Blakely⁵, David Peereboom⁶, Carol Kruse⁷, Walter Hall⁸, and Dennis Groothuis⁹

¹ Texas Tech University Health Sciences Center, Pharmaceutical Sciences, 1300 Coulter
Amarillo, Texas 79106, Quentin.Smith@ttuhsc.edu, 806-356-4016

² Memorial Sloan-Kettering Cancer Center, Neurology Department, 1275 York Avenue
New York, New York 10021, abreyl@mskcc.org, 212-639-5122

³ Massachusetts General Hospital, Boston, Massachusetts, USA

⁴ Cedars Sinai Medical Center, Los Angeles, California, USA

⁵ University of Manitoba, Winnipeg, Manitoba, Canada

⁶ Cleveland Clinic Foundation, Cleveland, Ohio, USA

⁷ La Jolla Institute for Molecular Medicine, San Diego, California, USA

⁸ SUNY Upstate Medical University, Syracuse, New York, USA

⁹ Northwestern University Medical School, Evanston, Illinois, USA

Introduction

The blood-brain barrier (BBB) is critical to the understanding and management of primary and metastatic brain tumors. This complex pathophysiologic structure has been extensively studied and manipulated in an effort to improve our understanding and management of brain tumors; however, many issues remain unresolved. Neuroimaging demonstrates clear disruption of the BBB in many tumors, yet available drug-based therapies are often excluded in whole or part from the tumor, possibly on the basis of efflux transporters and other restrictions. In this report we summarize the current state of the science and seek to address the areas which remain a barrier to progress and identify specific ways to move this field forward.

1. State of the Science - Progress Since 2000

Blood-Brain and Blood-Tumor Barriers

Since 2000, critical advances have been made at the cellular and molecular levels in the structure of the blood-brain barrier and how it changes with tumor invasion and growth. The transcriptome of the brain microvascular endothelium, which comprises the BBB, has been cataloged¹ and initial comparisons have been made between vascular endothelial cells in nonneoplastic and malignant brain tissue.² A number of proteins which comprise the endothelial tight junction, including claudin-3, -5 and -12, occludin, junctional adhesion molecule A and zonula occludens proteins, have been identified,³ and initial studies have been conducted examining changes induced by selective gene knockout.⁴ The brain microvascular endothelium has been shown to express a broad range of drug efflux transporter proteins that aggressively remove many chemotherapeutic drugs from the central nervous system. These transporters include ABCB1 (p-glycoprotein), ABCC1, 2, 4 and 5 (MRP), ABCG2 (BCRP), OAT-3, OATP1A2, and OATP2B1. Preliminary studies have demonstrated alterations in transporter expression in gliomas and the

blood-tumor barrier.⁵⁻⁷ The role of these transporters in limiting brain and brain tumor drug uptake has been established for a number of important chemotherapeutic agents (i.e., paclitaxel, imatinib, topotecan, vincristine, vinblastine, doxorubicin).⁸⁻¹⁰ Together, the low passive permeability of the BBB, coupled with active extrusion by transporter proteins, forms a potent barrier for brain delivery of many chemotherapeutic drug agents.

Multiple factors have been identified that modify vascular endothelial function and barrier tightness in pathologic conditions, including brain tumors.¹¹ Significant strides have been achieved in understanding the factors that control metastatic cell invasion of the nervous system as well as tumor implantation and angiogenesis.¹²⁻¹⁶ Novel imaging methods have also been introduced to map changes in *in vivo* blood-tumor barrier permeability to fluorescent probes¹⁷ or paramagnetic probes.¹⁸ Tumor seeding and growth in brain can be followed by MRI¹⁹ as well as luminescent methods^{20,21} that offer the ability to monitor the time course of drug efficacy and can be coupled with drug pharmacokinetics. Physiologic factors,²² including tumor interstitial fluid pressure, have been shown to markedly influence drug penetration to tumor. Lowering interstitial fluid pressure can enhance the time-concentration exposure of drug to tumor tissue.²³

Drug Delivery Strategies

A critical issue in the management of brain tumors is adequate delivery of chemotherapeutic agents to the tumor. This delivery may be impeded by the BBB (Figure 1), as well as osmotic pressure within the tumor itself. In order to better deliver drugs to brain tumors, multiple strategies to circumvent, disrupt or manipulate the BBB have been employed.

Convection enhanced delivery (CED), wherein chemotherapy is infused directly into the tumor via strategically placed catheters, has had significant progress from animal models to clinical trials in the last 10 years. CED is an attractive approach for agents that are thought to be too large to

cross the BBB or too toxic for systemic administration. This has been a particularly enticing approach for conjugated targeted toxin therapies and antibodies.^{24,25} The initial results with cintredekin besudotox (CB) delivered via CED showed safety and promising efficacy.^{26,27} However, the large phase 3 trial of CB versus carmustine impregnated wafers (PRECISE trial) was recently completed and did not meet its efficacy endpoint of a significant difference in the overall survival (OS). There was, however, activity with both CED of CB and carmustine impregnated wafers, resulting in overall survival rates of 14% at 24 months for recurrent or progressive GBM. Similarly, the TransMID (modified diphtheria toxin conjugated to transferrin) CED phase 3 trial was recently stopped at the interval analysis due to low probability of meeting efficacy endpoints. Final results from these studies are expected in the next year.

Challenges that may limit the success of CED are limited distribution of infused drug and selection of effective agents for infusion and neurotoxicities due to large volume infusions. To overcome these limitations, most clinical trials use ≥ 2 catheters for infusion to maximize distribution of drug to various regions of tumor. However, it is not yet clear that multiple catheters result in substantially improved distribution of drug throughout heterogeneous brain tumors. The ongoing phase II trial of chlorotoxin coupled with the radioisotope ^{131}I (^{131}I -TM-601) infuses agent into the tumor resection cavity via Ommaya reservoir.²⁸ This approach allows for repeat dosing, but remains limited in the potential volume of distribution. Reverse microdialysis in which drug is infused into tissue with small volumes has shown similar results to standard CED approaches, but may have even more difficulty achieving wide volumes of distribution. Recent studies have used liposomal drug preparations or solvent facilitated perfusion to increase the area of distribution of infused drug in preclinical and early clinical trials.²⁹ Increasingly, CED studies are employing imaging techniques such as FDG-PET, DWI MRI and SPECT to attempt to image the area of distribution of agent after infusion.³⁰ These techniques have not yet been validated with direct measures of drug concentration. However, this is an ongoing important area of research that may help in planning future trials to optimize CED and other local therapies.

Polymer-based local drug delivery to tumors has been part of standard care for select patients with malignant gliomas since the FDA approval of carmustine impregnated wafers for recurrent GBM in 1996. However, recent studies have shown that the maximal tolerated dose (MTD) of carmustine in polymers in patients with recurrent gliomas may in fact be 40 mg, rather than the 7.7mg dose currently approved and in clinical practice.³¹ Efficacy studies at the high dose are needed to determine if there is improved benefit, but such trials are difficult to efficiently advance due to the FDA requirement for approval of each polymer and drug combination. There has also been advancement in the biotechnology of local drug delivery with increasing variety of media being tested for local implantation. One such agent is ReGel, which is a viscous liquid when cooled, but becomes solid at room or body temperature, conforming to the shape of the cavity it occupies.³² Similar to polymers, it degrades over time with slow local release of the chemotherapeutic agent. Phase II trials using this technology with paclitaxel for patients with recurrent gliomas are now being planned. Other local delivery techniques, such as cisplatin-infused plates placed into tumor, have also shown initial promising results.³³ The benefits of these approaches are direct drug delivery to the tumor with minimal systemic or local toxicities and little added complexity or risk to surgical procedures. Similar to CED, limitations include uneven distribution of the drug to heterogeneous infiltrating tumors as well as variable release of the chemotherapeutic agent.

Blood-brain barrier disruption (BBBD), with or without intra-arterial (IA) chemotherapy, has been extensively investigated as a technique to permit agents to go through, rather than around, the BBB. Most commonly this approach utilizes osmotic BBBD with agents such as mannitol followed by IA chemotherapy. However, more recently, MRI-guided ultrasound-induced BBBD has been investigated.^{34,35} Although relatively small numbers of patients have been treated, there have been promising results with BBBD and IA chemotherapy including in traditionally difficult tumors such as brainstem gliomas.³⁶ There also may be particular promise for long-term complete responses without the cognitive loss due to whole brain radiation therapy³⁷ when this technique is used with methotrexate based therapy for primary central nervous system lymphoma

(PCNSL). Delivery of antibodies such as Rituxan, is more difficult (Figure 2), than smaller molecules such as methotrexate³⁸ but now can be studied in PCNSL animal models.³⁹

MRI techniques are increasingly being used with this approach in an attempt to quantify the degree of BBBD achieved. Interestingly, standard therapy such as conformal radiation therapy (XRT) has been demonstrated to alter the BBB permeability to gadolinium diethylenetriamine-pentaacetic acid (Gd-DTPA) in patients with high grade gliomas. Initially, non-enhancing tumor had significantly increased contrast uptake after 30 Gy, whereas initially contrast-enhancing tumor had a decrease in degree of contrast enhancement over time. The results suggest that XRT alone may induce BBBD potentially allowing increased entry of anti-cancer agents. This is particularly notable given the recent improvement in survival in GBM with the regimen of temozolomide given concurrently with XRT.

Modifying systemic agents to increase their permeability or enhance receptor mediated transport across the BBB is of increasing interest. Technological advances in the delivery of drugs via nanoparticles also continue to develop. Nanoparticles are particularly attractive as they can be engineered to maximize transport across the BBB and can be “loaded” with both therapeutic agents and agents to optimize various neuroimaging techniques.⁴⁰⁻⁴⁴ Currently, this technology remains in pre-clinical development.

Tumor and Brain Drug Pharmacokinetics

Key questions for any delivery approach are what amount of drug is delivered and to which area of tumor. It is becoming increasingly apparent that determining intratumoral drug concentrations and local effects of investigational agents is an important step in early phase clinical trials. There are multiple techniques in development for assessing drug delivery to brain tumors.

Microdialysis uses small, flexible catheters with specialized permeable membranes placed within tumor to sample the extracellular fluid. This has been used extensively in preclinical trials to assess drug PK within tumor and can be used to measure brain concentrations after both systemic and local delivery. The first clinical trial to measure intratumoral drug concentrations in brain cancer patients with microdialysis catheters was recently completed through the New Approaches to Brain Tumor Therapy (NABTT) consortium.⁴⁵ Results demonstrated: 1) the feasibility of the technique for obtaining continuous intratumoral pharmacokinetic data in glioma patients, and 2) regional variability in tumor drug concentration relative to BBB integrity. Similar feasibility has been shown using this technique to measure p-boronophenylate in tumors for glioma patients undergoing boron neutron capture therapy.⁴⁶ Additional studies in brain cancer patients to measure differential concentrations of investigational agents within tumor are currently under way. This technique is valuable in early phase clinical trials not only to determine investigational drug PK, but also to assess local effects of the agent. Such information may be particularly meaningful for pharmacodynamic determinations of optimal doses of small molecular-based therapies that are unlikely to have the toxicities on which traditional MTD studies depend. Although microdialysis is a powerful and readily available tool, it is limited by regional sampling of the extracellular fluid, requiring multiple catheters to assess heterogeneous portions of tumors, inability to sample substances > 20kD, and technical considerations regarding recovery.

Another approach to determining intratumoral drug concentration and effect is tissue sampling. This approach is preferable for assessment of large molecules and membrane bound proteins such as receptors. An increasingly popular clinical trial design is to treat patients with the investigational agent for some period prior to a planned resection and then sample the tumor at the time of resection for drug concentration and targeted effect.⁴⁷ The limitations of this approach are that it requires the patient have surgery and gives only one time point regarding drug concentration and activity, The trauma of surgery can also impact blood-tumor and blood- brain pharmacology. Accurate MR assessment of tumor blood volume, and flow without the artifact of

gadolinium leakage requires blood pool agents such as nanoparticles⁴⁸ to complement changes in permeability using gadolinium following anti- angiogenesis therapy

Imaging

Brain imaging technologies have been applied with increasing success to measure the delivery and local effects of anti-cancer agents in brain tumors. Batchelor et al. recently applied a variety of MRI sequences for quantification of tumor vessel response to a systemic pan-VEGF receptor tyrosine kinase inhibitor (AZD2171) in patients with recurrent GBM.⁴⁹ They were able to show that with antiangiogenic therapy, there was progressive normalization of microvessel size, volume of contrast enhancement and edema. Moreover, the MRI measures were correlated with circulating markers of angiogenesis, such as FGF2 and SDF1, as well as viable circulating endothelial cells. Although this approach does not give specific intratumoral PK data, it does show the activity of the agent non-invasively over time, providing a new technique for serial monitoring of drug effect on the tumor.

Techniques to quantify cerebral blood volume are under intense investigation and may add additional sensitivity to existing imaging sequences for evaluation of tumor vasculature in response to various interventions. Investigation of molecular imaging agents is also under intense development for important tumor targets such as EGFR. MRI can also be used to measure delivery of drug to tumor directly, particularly with agents known to shorten T1 such as the radiation enhancing agent motexafin gadolinium. Single photon emission computed tomography (SPECT) has been increasingly used in clinical trials to measure delivery of labeled drug brain. Interestingly, SPECT has also been successfully applied in conjunction with PET to assess the activity of multidrug resistance (MDR1) P-glycoprotein (Pgp) in cell culture, suggesting that non-invasive monitoring of key BBB transporters in response to various chemotherapeutics may be possible *in vivo*.

It is critical to note that delivering drug directly to tumor is often associated with pathologic imaging changes that may confound assessments of tumor progression. This has been demonstrated for CED, polymer-based therapies, immunotherapies, and BBBD approaches. In particular, immune therapies can increase inflammatory events dramatically. A flare phenomenon has been observed in MRIs performed post-immunotherapy. Interpretation of increased mass effect and edema have resulted in inadvertent placement of patients off protocol. Better discernment of tumor progression and inflammation are needed. Novel applications of PET, SPECT and new MRI sequences are being investigated to better distinguish tumor growth from treatment-induced injury. These imaging changes remain a complicating factor in the clinical management of brain tumors after drug delivery across the BBB.

In an NIH-sponsored conference, “a reality check” of tumor delivery, such as those discussed above, has been published.⁵⁰

2. Barriers to Progress

Brain tumors are heterogeneous both genotypically and phenotypically. The BBB-brain tumor interface may be similar or quite different in different types of tumors. Current *in vitro* and animal models are inadequate for studying the BBB-brain tumor interface. Brain metastases are by far the most common brain tumor and yet the least studied. By sheer number it makes sense to focus on brain metastases which may provide unique insights into the molecular pathogenesis of tumors that home to the brain.

There is a lack of information regarding pharmacokinetic and pharmacodynamic endpoints in the brain. Most studies to date have largely focused on whether or not drug can be detected in tumor or even CSF as a surrogate for brain/brain tumor concentration. It will be critical to consider issues such as effective target tissue concentrations, dose and duration of exposure, timing of

measurements, receptor saturation and the variability of drug levels at different regions within the tumor. The ability to directly measure drug delivery to tumors is difficult. Current techniques, such as tissue drug levels and microdialysate drug levels, require surgical intervention and demand intensive time and effort on the part of a multidisciplinary team to coordinate. Small variations, such as the location of catheter placement, can have a significant impact on results. Further research is needed to define the efflux transporters and their activity as associated with various antitumor agents. Surrogate measures of the BBB-brain tumor interface, such as imaging surrogates, are difficult to validate and are often expensive.

There are interdisciplinary barriers to advancing the understanding of BBB-brain tumor interface. The groups that need to be able to interact include neurosurgeons, neurologists, neuro-oncologists, medical oncologists, neurobiologists, neuroradiologists and pharmacologists. This is a barrier in establishing effective collaborations and communicating existing data.

Interdisciplinary grants may have a difficult time in the review process for a number of reasons including standard committee review criteria or being submitted to the wrong institute/program. This is exacerbated by an oversimplified understanding or appreciation of the importance and role of the BBB by many professions involved in the care of brain tumor patients.

Although a number of different strategies have been developed to technically improve the delivery of various agents to brain tumors, phase 3 trials, albeit difficult, are needed to show incremental therapeutic benefit for the patient. On the other hand, increasing delivery as reviewed by a statistician appears to impact efficacy.⁵¹ An important goal moving forward would be to target both effective delivery strategies and to pair these with agents that have the most promise if delivered efficiently. The expense and regulatory requirements related to the conduct of clinical trials is a significant barrier to progress.

3. Specific Recommendations to Advance the Field

Key Basic Science Research areas:

- Develop representative tumor model systems that can be used in animals to monitor tumor growth and drug action noninvasively.
- Characterize the proteome and genome of the BBB-brain tumor interfaces and how they change with tumor progression and treatment. Included in this would be factors that influence tumor cell metastasis and invasion of brain, including growth and angiogenesis.
- The factors that determine drug transport across the BBB so that “brain” or “brain tumor” available drugs can be designed de novo.
- The role of tumor invasion and micrometastases, particular those tumors which develop in the setting of an intact BBB.
- Delivery of targeted molecules, including large molecular weight antibodies to treat primary and metastatic CNS tumors.

Key Clinical Research areas:

- Develop surrogate measures of the BBB-brain tumor interface (including permeability, drug delivery, etc). In particular, new imaging modalities are likely to be critical and it will be important to encourage interactions with expert radiologists and the imaging branch of the National Institutes of Health to optimize these modalities. However, it is also felt to be critical that these imaging surrogates be validated or confirmed with actual tissue assays.
- Develop nanotechnology-based drug delivery, receptor-mediated drug delivery, and carrier-mediated drug delivery to target and enhance drug uptake into tumors and surrounding brain tissue.
- Develop Phase 0 clinical trials to optimize drug and tissue assays appropriate to validate the delivery of novel agents or novel delivery strategies before proceeding to larger more traditional trials.
- Future clinical trials should include:
 - Tissue studies of major efflux transporters in newly diagnosed patients
 - Brain metastasis

- Pre-irradiation studies
- Microdialysis studies in low grade-gliomas where the BBB is presumably intact could be valuable
- Drugs that inhibit the major efflux transport systems

Education

- To educate and integrate medical professionals involved in brain tumor research at all levels about the importance of the BBB. This includes but is not limited to clinicians caring for brain tumor patients, pharmaceutical companies and laboratories.
- Lobby American Society of Clinical Oncology or Society of Neuro-Oncology to dedicate an education session to the relevance of the BBB in the understanding and treatment of metastatic and primary brain tumors.

Specific Resources

- Funding initiatives targeted to BBB and blood tumor barrier research. These initiatives should clearly emphasize and truly support interdisciplinary research. This funding is critical both for the basic science as well as the clinical research areas.
- In addition to specific research funding, it is critical to have adequate insurance coverage for standard procedures to allow the conduct of clinical trials.

4. Single Most Important Issue

Multiple topics were discussed during the meeting that are of importance to brain barrier systems and tumors. The consensus of the group at the meeting was that imaging was the single most important issue. Currently, *in vivo* imaging of brain tumors is limited by a number of factors. The skull and current methods restrict the accurate assessment of tumor size, location and type as

well as response to therapy. In most studies, information obtained is constrained to a small subset of properties, such as barrier passive permeability, blood volume, blood flow, and glucose metabolism. Improved imaging techniques, which offer the opportunity to accurately monitor tumor invasion, growth, angiogenesis, cellular apoptosis, necrosis, inflammation, edema, and gene expression, would greatly aid studies of brain tumor biology. Such may be combined with continuous assays of tumor drug delivery to more appropriately assess *in vivo* response to therapy. A need exists for quantitative imaging methods that can be used in humans and small experimental animals to characterize tumors and their response to therapeutic interventions.

References

1. Enerson BE, Drewes LR. The rat blood-brain barrier transcriptome. *J Cereb Blood Flow Metab* 2006;**26**(7):959-73.
2. Madden SL, Cook BP, Nacht M, Weber WD, Callahan MR, Jiang Y, Dufault MR, Zhang X, Zhang W, Walter-Yohrling J, Rouleau C, Akmaev VR, Wang CJ, Cao X, St Martin TB, Roberts BL, Teicher BA, Klinger KW, Stan RV, Lucey B, Carson-Walter EB, Laterra J, Walter KA. Vascular gene expression in nonneoplastic and malignant brain. *Am J Pathol* 2004;**165**(2):601-8.
3. Hawkins BT, Davis TP. The blood-brain barrier/neurovascular unit in health and disease. *Pharmacol Rev* 2005;**57**(2):173-85.
4. Nitta T, Hata M, Gotoh S, Seo Y, Sasaki H, Hashimoto N, Furuse M, Tsukita S. Size-selective loosening of the blood-brain barrier in claudin-5-deficient mice. *J Cell Biol* 2003;**161**(3):653-60.
5. Regina A, Demeule M, Laplante A, Jodoin J, Dagenais C, Berthelet F, Moghrabi A, Beliveau R. Multidrug resistance in brain tumors: roles of the blood-brain barrier. *Cancer Metastasis Rev* 2001;**20**(1-2):13-25.
6. Kushihara H, Sugiyama Y. Active efflux across the blood-brain barrier: role of the solute carrier family. *NeuroRx* 2005;**2**(1):73-85.
7. Bronger H, Konig J, Kopplow K, Steiner HH, Ahmadi R, Herold-Mende C, Keppler D, Nies AT. ABC drug efflux pumps and organic anion uptake transporters in human gliomas and the blood-tumor barrier. *Cancer Res* 2005;**65**(24):11419-28.
8. Fellner S, Bauer B, Miller DS, Schaffrik M, Fankhanel M, Spruss T, Bernhardt G, Graeff C, Farber L, Gschaidmeier H, Buschauer A, Fricker G. Transport of paclitaxel (Taxol) across the blood-brain barrier in vitro and in vivo. *J Clin Invest* 2002;**110**(9):1309-18.

9. Leggas M, Adachi M, Scheffer GL, Sun D, Wielinga P, Du G, Mercer KE, Zhuang Y, Panetta JC, Johnston B, Scheper RJ, Stewart CF, Schuetz JD. Mrp4 confers resistance to topotecan and protects the brain from chemotherapy. *Mol Cell Biol* 2004;**24**(17):7612-21.
10. Breedveld P, Beijnen JH, Schellens JH. Use of P-glycoprotein and BCRP inhibitors to improve oral bioavailability and CNS penetration of anticancer drugs. *Trends Pharmacol Sci* 2006;**27**(1):17-24.
11. Abbott NJ, Ronnback L, Hansson E. Astrocyte-endothelial interactions at the blood-brain barrier. *Nat Rev Neurosci* 2006;**7**(1):41-53.
12. Lee TH, Avraham HK, Jiang S, Avraham S. Vascular endothelial growth factor modulates the transendothelial migration of MDA-MB-231 breast cancer cells through regulation of brain microvascular endothelial cell permeability. *J Biol Chem* 2003;**278**(7):5277-84.
13. Lee BC, Lee TH, Avraham S, Avraham HK. Involvement of the chemokine receptor CXCR4 and its ligand stromal cell-derived factor 1alpha in breast cancer cell migration through human brain microvascular endothelial cells. *Mol Cancer Res* 2004;**2**(6):327-38.
14. Avraham HK, Lee TH, Koh Y, Kim TA, Jiang S, Sussman M, Samarel AM, Avraham S. Vascular endothelial growth factor regulates focal adhesion assembly in human brain microvascular endothelial cells through activation of the focal adhesion kinase and related adhesion focal tyrosine kinase. *J Biol Chem* 2003;**278**(38):36661-8.
15. Steeg PS. Tumor metastasis: mechanistic insights and clinical challenges. *Nat Med* 2006;**12**(8):895-904.
16. Naumov GN, Akslen LA, Folkman J. Role of angiogenesis in human tumor dormancy: animal models of the angiogenic switch. *Cell Cycle* 2006;**5**(16):1779-87.

17. Lu W, Bucana CD, Schroit AJ. Pathogenesis and vascular integrity of breast cancer brain metastasis. *Int J Cancer* 2007;**120**(5):1023-6.
18. Ewing JR, Brown SL, Lu M, Panda S, Ding G, Knight RA, Cao Y, Jiang Q, Nagaraja TN, Churchman JL, Fenstermacher JD. Model selection in magnetic resonance imaging measurements of vascular permeability: Gadomer in a 9L model of rat cerebral tumor. *J Cereb Blood Flow Metab* 2006;**26**(3):310-20.
19. Heyn C, Ronald JA, Ramadan SS, Snir JA, Barry AM, MacKenzie LT, Mikulis DJ, Palmieri D, Bronder JL, Steeg PS, Yoneda T, MacDonald IC, Chambers AF, Rutt BK, Foster PJ. In vivo MRI of cancer cell fate at the single-cell level in a mouse model of breast cancer metastasis to the brain. *Magn Reson Med* 2006;**56**(5):1001-10.
20. Kemper EM, Leenders W, Kusters B, Lyons S, Buckle T, Heerschap A, Boogerd W, Beijnen JH, van Tellingen O. Development of luciferase tagged brain tumour models in mice for chemotherapy intervention studies. *Eur J Cancer* 2006;**42**(18):3294-303.
21. Szentirmai O, Baker CH, Lin N, Szucs S, Takahashi M, Kiryu S, Kung AL, Mulligan RC, Carter BS. Noninvasive bioluminescence imaging of luciferase expressing intracranial U87 xenografts: correlation with magnetic resonance imaging determined tumor volume and longitudinal use in assessing tumor growth and antiangiogenic treatment effect. *Neurosurgery* 2006;**58**(2):365-72; discussion 365-72.
22. Minchinton AI, Tannock IF. Drug penetration in solid tumours. *Nat Rev Cancer* 2006;**6**(8):583-92.
23. Navalitloha Y, Schwartz ES, Groothuis EN, Allen CV, Levy RM, Groothuis DR. Therapeutic implications of tumor interstitial fluid pressure in subcutaneous RG-2 tumors. *Neuro-oncol* 2006;**8**(3):227-33.
24. Sampson JH, Akabani G, Archer GE, Bigner DD, Berger MS, Friedman AH, Friedman HS, Herndon JE, 2nd, Kunwar S, Marcus S, McLendon RE, Paolino A, Penne K,

- Provenzale J, Quinn J, Reardon DA, Rich J, Stenzel T, Tourt-Uhlig S, Wikstrand C, Wong T, Williams R, Yuan F, Zalutsky MR, Pastan I. Progress report of a Phase I study of the intracerebral microinfusion of a recombinant chimeric protein composed of transforming growth factor (TGF)-alpha and a mutated form of the Pseudomonas exotoxin termed PE-38 (TP-38) for the treatment of malignant brain tumors. *J Neurooncol* 2003;**65**(1):27-35.
25. Patel SJ, Shapiro WR, Laske DW, Jensen RL, Asher AL, Wessels BW, Carpenter SP, Shan JS. Safety and feasibility of convection-enhanced delivery of Cotara for the treatment of malignant glioma: initial experience in 51 patients. *Neurosurgery* 2005;**56**(6):1243-52; discussion 1252-3.
26. Kunwar S, Chang SM, Prados MD, Berger MS, Sampson JH, Croteau D, Sherman JW, Grahn AY, Shu VS, Dul JL, Husain SR, Joshi BH, Pedain C, Puri RK. Safety of intraparenchymal convection-enhanced delivery of cintredekin besudotox in early-phase studies. *Neurosurg Focus* 2006;**20**(4):E15.
27. Weber F, Asher A, Bucholz R, Berger M, Prados M, Chang S, Bruce J, Hall W, Rainov NG, Westphal M, Warnick RE, Rand RW, Floeth F, Rommel F, Pan H, Hingorani VN, Puri RK. Safety, tolerability, and tumor response of IL4-Pseudomonas exotoxin (NBI-3001) in patients with recurrent malignant glioma. *J Neurooncol* 2003;**64**(1-2):125-37.
28. Mamelak AN, Rosenfeld S, Bucholz R, Raubitschek A, Nabors LB, Fiveash JB, Shen S, Khazaeli MB, Colcher D, Liu A, Osman M, Guthrie B, Schade-Bijur S, Hablitz DM, Alvarez VL, Gonda MA. Phase I single-dose study of intracavitary-administered iodine-131-TM-601 in adults with recurrent high-grade glioma. *J Clin Oncol* 2006;**24**(22):3644-50.
29. Hamstra DA, Moffat BA, Hall DE, Young JM, Desmond TJ, Carter J, Pietronigro D, Frey KA, Rehemtulla A, Ross BD. Intratumoral injection of BCNU in ethanol (DTI-015) results in enhanced delivery to tumor--a pharmacokinetic study. *J Neurooncol* 2005;**73**(3):225-38.

30. Voges J, Reszka R, Gossmann A, Dittmar C, Richter R, Garlip G, Kracht L, Coenen HH, Sturm V, Wienhard K, Heiss WD, Jacobs AH. Imaging-guided convection-enhanced delivery and gene therapy of glioblastoma. *Ann Neurol* 2003;**54**(4):479-87.
31. Olivi A, Grossman SA, Tatter S, Barker F, Judy K, Olsen J, Bruce J, Hilt D, Fisher J, Piantadosi S. Dose escalation of carmustine in surgically implanted polymers in patients with recurrent malignant glioma: a New Approaches to Brain Tumor Therapy CNS Consortium trial. *J Clin Oncol* 2003;**21**(9):1845-9.
32. Vukelja SJ, Anthony SP, Arseneau JC, Berman BS, Cunningham CC, Nemunaitis JJ, Samlowski WE, Fowers KD. Phase 1 study of escalating-dose OncoGel (ReGel/paclitaxel) depot injection, a controlled-release formulation of paclitaxel, for local management of superficial solid tumor lesions. *Anticancer Drugs* 2007;**18**(3):283-9.
33. Sheleg SV, Korotkevich EA, Zhavrid EA, Muravskaya GV, Smeyanovich AF, Shanko YG, Yurkshtovich TL, Bychkovsky PB, Belyaev SA. Local chemotherapy with cisplatin-depot for glioblastoma multiforme. *J Neurooncol* 2002;**60**(1):53-9.
34. Kinoshita M, McDannold N, Jolesz FA, Hynynen K. Noninvasive localized delivery of Herceptin to the mouse brain by MRI-guided focused ultrasound-induced blood-brain barrier disruption. *Proc Natl Acad Sci U S A* 2006;**103**(31):11719-23.
35. Hynynen K, McDannold N, Vykhodtseva N, Jolesz FA. Noninvasive MR imaging-guided focal opening of the blood-brain barrier in rabbits. *Radiology* 2001;**220**(3):640-6.
36. Hall WA, Doolittle ND, Daman M, Bruns PK, Muldoon L, Fortin D, Neuwelt EA. Osmotic blood-brain barrier disruption chemotherapy for diffuse pontine gliomas. *J Neurooncol* 2006;**77**(3):279-84.
37. Jahnke K, Doolittle ND, Muldoon LL, Neuwelt EA. Implications of the blood-brain barrier in primary central nervous system lymphoma. *Neurosurg Focus* 2006;**21**(5):E11.

38. Doolittle ND, Jahnke K, Belanger R, Ryan DA, Nance RWJ, Lacy CA, Tyson RM, Haluska M, Hedrick NA, varallyay C, Neuwelt EA. Potential of immunotherapy and radioimmunotherapy in relapsed primary CNS lymphoma. *Leukemia and Lymphoma* 2007;**48**(9): 1712-1720.
39. Soussain C, Muldoon LL, Varallyay C, Jahnke K, DePaula L, Neuwelt EA. Characterization and magnetic resonance imaging of a rat model of human B-cell central nervous system lymphoma. *Clin Cancer Res* 2007;**13**(8):2504-11.
40. Petri B, Bootz A, Khalansky A, Hekmatara T, Muller R, Uhl R, Kreuter J, Gelperina S. Chemotherapy of brain tumour using doxorubicin bound to surfactant-coated poly(butyl cyanoacrylate) nanoparticles: revisiting the role of surfactants. *J Control Release* 2007;**117**(1):51-8.
41. Kohler N, Sun C, Fichtenholtz A, Gunn J, Fang C, Zhang M. Methotrexate-immobilized poly(ethylene glycol) magnetic nanoparticles for MR imaging and drug delivery. *Small* 2006;**2**(6):785-92.
42. Reddy GR, Bhojani MS, McConville P, Moody J, Moffat BA, Hall DE, Kim G, Koo YE, Woolliscroft MJ, Sugai JV, Johnson TD, Philbert MA, Kopelman R, Rehemtulla A, Ross BD. Vascular targeted nanoparticles for imaging and treatment of brain tumors. *Clin Cancer Res* 2006;**12**(22):6677-86.
43. u W, Sun Q, Wan J, She Z, Jiang XG. Cationic albumin-conjugated pegylated nanoparticles allow gene delivery into brain tumors via intravenous administration. *Cancer Res* 2006;**66**(24):11878-87.
44. Ambruosi A, Khalansky AS, Yamamoto H, Gelperina SE, Begley DJ, Kreuter J. Biodistribution of polysorbate 80-coated doxorubicin-loaded [¹⁴C]-poly(butyl cyanoacrylate) nanoparticles after intravenous administration to glioblastoma-bearing rats. *J Drug Target* 2006;**14**(2):97-105.

45. Olson JJ, Blakeley JO, Grossman SA, Weingart J, Rashid A, Supko J, Nabtt CNSC. Differences in the distribution of methotrexate into high grade gliomas following intravenous administration, as monitored by microdialysis, are associated with blood brain barrier integrity. *J Clin Oncol (Meeting Abstracts)* 2006;**24**(18_suppl):1548.
46. Bergenheim AT, Capala J, Roslin M, Henriksson R. Distribution of BPA and metabolic assessment in glioblastoma patients during BNCT treatment: a microdialysis study. *J Neurooncol* 2005;**71**(3):287-93.
47. Lassman AB, Rossi MR, Raizer JJ, Abrey LE, Lieberman FS, Grefe CN, Lamborn K, Pao W, Shih AH, Kuhn JG, Wilson R, Nowak NJ, Cowell JK, DeAngelis LM, Wen P, Gilbert MR, Chang S, Yung WA, Prados M, Holland EC. Molecular study of malignant gliomas treated with epidermal growth factor receptor inhibitors: tissue analysis from North American Brain Tumor Consortium Trials 01-03 and 00-01. *Clin Cancer Res* 2005;**11**(21):7841-50.
48. Neuwelt EA, Varallyay CG, Manninger S, Solymosi D, Haluska M, Hunt MA, Nesbit G, Stevens A, Jerosch-Herold M, Jacobs PM, Hoffman JM. The potential of ferumoxytol nanoparticle magnetic resonance imaging, perfusion, and angiography in central nervous system malignancy: a pilot study. *Neurosurgery* 2007;**60**(4):601-11; discussion 611-2.
49. Batchelor TT, Sorensen AG, di Tomaso E, Zhang WT, Duda DG, Cohen KS, Kozak KR, Cahill DP, Chen PJ, Zhu M, Ancukiewicz M, Mrugala MM, Plotkin S, Drappatz J, Louis DN, Ivy P, Scadden DT, Benner T, Loeffler JS, Wen PY, Jain RK. AZD2171, a pan-VEGF receptor tyrosine kinase inhibitor, normalizes tumor vasculature and alleviates edema in glioblastoma patients. *Cancer Cell* 2007;**11**(1):83-95.
50. Muldoon L, Soussain C, Jahnke K, Johanson C, Siegal T, Smith QR, Hall W, Hynynen K, Senter PD, Peereboom DM, Neuwelt EA. Chemotherapy Delivery Issues in Central Nervous System Malignancy: A Reality Check. *J Clin Oncol* 2007: **25**(16): 2295-2305.

51. Kraemer DF, Fortin D, Doolittle ND, Neuwelt EA. Association of total dose intensity of chemotherapy in primary central nervous system lymphoma (human non-acquired immunodeficiency syndrome) and survival. *Neurosurgery* 2001;**48**(5):1033-40; discussion 1040-1.
52. Smith QR. A review of blood-brain barrier transport techniques. *Methods Mol Med* 2003;**89**:193-208.

Figure Legends

Figure 1. Relationship between BBB permeability and octanol/water partition coefficient for chemotherapeutic agents. The solid line is the least-squares fit to the data for agents that are not actively taken up by brain or pumped out by the BBB (adapted from Smith, 2003) The permeability to specific chemotherapeutic agents is shown.⁵² Many chemotherapeutics show limited uptake into brain because they are either polar (low octanol/water distribution coefficient), have molecular weight >500, bind extensively to plasma proteins, or are transported out of brain by active efflux pumps.

Figure 2.

Ibritumomab (Zevalin™) delivery and efficacy in PCNSL. Panels A and B: SPECT images at 24 hours (A) or 120 hours (B) after IV administration of 5.2 mCi ¹¹¹In-ibritumomab in a single case study. Panel B shows questionable uptake at the arrow. At 45 hours after ¹¹¹In-ibritumomab administration, decay-corrected radioactivity was determined in CSF (1579 dpm) and serum (572,443 dpm), giving a CSF/plasma ratio of 0.0028. Panels C-E: Magnetic resonance imaging of tumor response in the same case study. T1-weighted with gadolinium contrast axial scans are shown for (C) Pre-⁹⁰Y-ibritumomab showing enhancing tumor (i.e. high signal) around the occipital horn of the ventricle; (D) complete response 2 months after ⁹⁰Y-ibritumomab; (E) Relapse around the opposite occipital horn 3 months after ⁹⁰Y-ibritumomab, with continuing complete response at the site of the original tumor. Figure previously published in the Journal of Clinical Oncology.⁵⁰

Figure 1

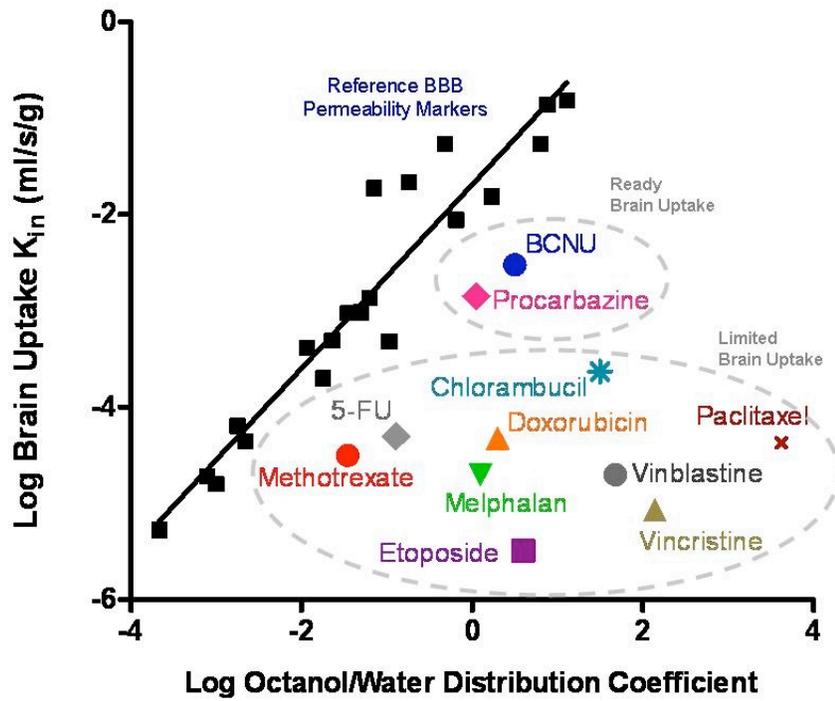


Figure 2

